

# Signaling Complex Organization by PDZ Domain Proteins

Jing-Song Fan Mingjie Zhang

Department of Biochemistry, Hong Kong University of Science and Technology, Kowloon, Hong Kong, PR China

## Key Words

PDZ domains · Scaffolding · Signal transduction · Domain organization

## Abstract

As one of the most abundant protein domains in the genomes of metazoans, PDZ domains play important roles in the targeting of proteins to specific cell membranes, as well as assembling proteins into supramolecular signaling complexes. The structures of individual PDZ domains, along with their diverse cooccurrence with a great variety of other protein domains, provide the biochemical basis for the functional diversity of PDZ proteins. In this review, we first briefly summarize the structure and target-binding properties of PDZ domains. After surveying the SMART protein domain database, we attempt to classify PDZ domain proteins into three general categories. We end the review by presenting several recent studies showing some novel features of PDZ domain proteins.

Copyright © 2003 S. Karger AG, Basel

PDZ domains are globular protein interaction modules that play central roles in organizing signal transduction complexes and specialized membrane structures. These domains were first identified from consensus sequences of ~90 amino acid residues of three proteins: the

postsynaptic density PSD-95/SAP90, the *Drosophila* septate junction protein disc-large, and the tight junction protein ZO-1. PDZ domains were also referred to as DHR (DLG homology region) or the GLGF repeat (after a signature Gly-Leu-Gly-Phe sequence in the domain) at an early stage in the literature. Today, it is known that PDZ domains are among the most abundant protein domains in the multicellular eukaryotic genomes. Analysis of the genome sequences of *Caenorhabditis elegans*, *Drosophila* and humans estimates the presence of 89, 128, and 320 PDZ domain-containing proteins, respectively. The number of PDZ domains is significantly larger than these numbers in each genome, as many PDZ proteins contain multiple PDZ domains (see below) [1, 2]. Although the majority of PDZ proteins do not contain transmembrane domains, PDZ proteins are often associated with the cellular membrane. This association with the cell membrane enables PDZ proteins to organize localized protein complexes for signal transduction purposes [3–5]. Another general function of PDZ proteins is to organize/maintain large and complicated cellular structures such as cell-cell junctions [6, 7].

## Structures and Target Binding of PDZ Domains

The topic of the structural and target-binding properties of PDZ domains was covered in a number of recent excellent reviews [5, 8–11], and we will only provide a

## KARGER

Fax +41 61 306 12 34  
E-Mail karger@karger.ch  
www.karger.com

© 2003 S. Karger AG, Basel  
1424–862X/02/0116–0315\$19.50/0

Accessible online at:  
www.karger.com/nsg

Mingjie Zhang  
Department of Biochemistry, Hong Kong University of Science and Technology  
Clear Water Bay  
Kowloon, Hong Kong (PR China)  
Tel. +86 852 2358 8709, Fax +86 852 2358 1552, E-Mail mzhang@ust.hk

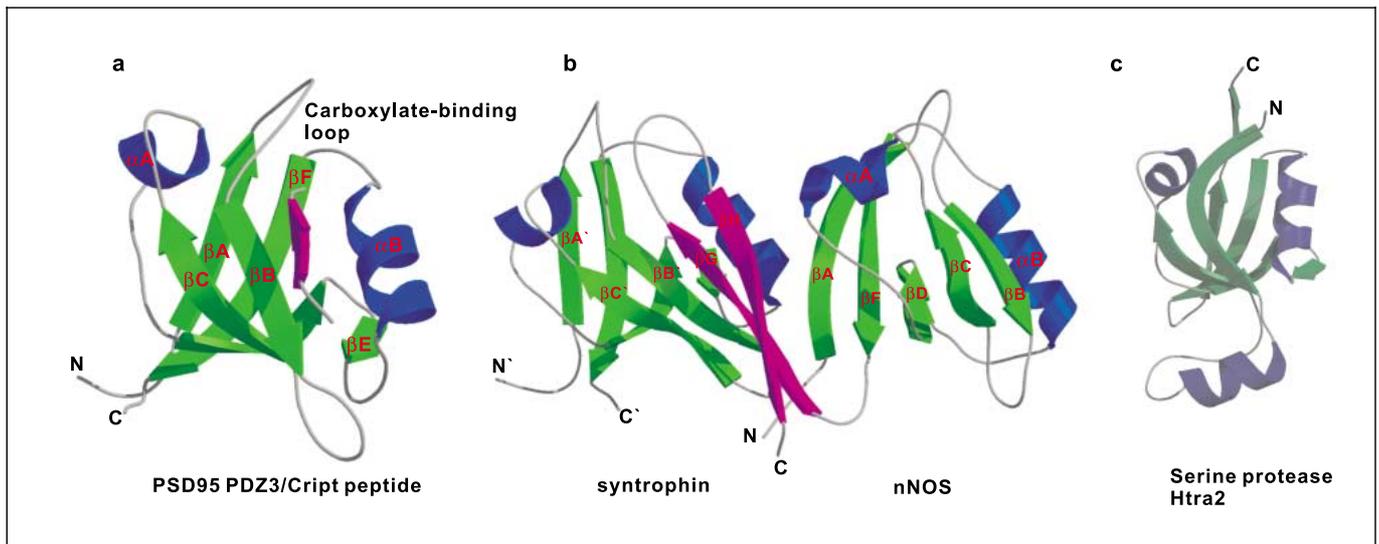


Fig. 1. Three-dimensional structures of PDZ domains in ribbon diagram representations. a The crystal structure of the third PDZ domain of PSD-95 in complex with a target peptide (in magenta) from the protein CRIPT [13], showing the structure and carboxyl peptide binding of a canonical PDZ domain. b The three-dimensional structure of the syntrophin/nNOS PDZ dimer, showing the binding of a PDZ domain to an internal peptide sequence [17]. The structure also highlights the mechanism of head-to-tail, PDZ domain-mediated dimerization. c The structure of the PDZ domain from mitochondrial serine protease HtrA2, showing the structure of an unconventional PDZ domain [47]. Note that the structure of the HtrA2 PDZ domain is permuted relative to the PDZ domains shown in a and b.

brief summary of this area. The structures of a number of canonical PDZ domains, both in free and target-bound forms, have been determined by X-ray crystallographic and/or NMR spectroscopic techniques [12–25]. PDZ domains fold into compact globular structure comprising a six-stranded  $\beta$ -barrel flanked by two  $\alpha$ -helices (fig. 1a). Peptide ligands from the extreme carboxyl termini of target proteins bind to a groove formed by the second  $\alpha$ -helix ( $\alpha$ B) and the second  $\beta$ -strand ( $\beta$ B) of the PDZ domains. The carboxyl peptides augment the  $\beta$ B strand in an anti-parallel fashion (fig. 1a). Amino acid residues at the 0 and –2 positions of the carboxyl peptide play dominant roles in the peptide's binding to a cognate PDZ domain, although residues at the –1 and –3 positions and those at further upstream positions also contribute to the binding [22, 23, 26, 27]. The uneven distribution of PDZ domain proteins as well as their binding partners provides a spatial regulation to the binding specificity of the interactions *in vivo*.

In addition to binding to carboxyl peptides, PDZ domains can also interact with internal short peptide fragments in target proteins [16, 17, 28–30]. The best example of this type of interaction is heterodimer formation be-

tween PDZ domains of nNOS and PSD-95 or syntrophin [16, 17, 29, 30]. The nNOS PDZ domain contains an additional two-stranded  $\beta$ -sheet (referred to as  $\beta$ -finger) at the C-terminal end of the canonical PDZ domain, and this  $\beta$ -finger structure inserts into the  $\alpha$ B/ $\beta$ B groove of the syntrophin/PSD-95 PDZ domain [17, 30] (fig. 1b). Mutational studies showed that a preformed  $\beta$ -finger structure is essential for the nNOS PDZ domain to bind to PSD-95/syntrophin [30, 31]. It is likely that the interaction between a PDZ domain and an internal peptide sequence depended on the specific amino acid sequence as well as the defined conformation of the peptide fragment. Due to the high stringency, the interaction between PDZ domain and internal sequences is likely to be less common. The studies of the interaction between nNOS and PSD-95/syntrophin provide a mechanistic understanding of PDZ domain-mediated oligomerization of PDZ domain proteins. However, it remains to be tested whether such a head-to-tail PDZ domain assembly is a general mechanism for PDZ domain-mediated oligomerization observed in a number of PDZ domain proteins [32–36].

## Domain Organization of PDZ-Containing Proteins

Inspection of ~1,250 PDZ proteins documented in the SMART database (<http://SMART.embl-heidelberg.de>) reveals that PDZ domains are organized in a variety of ways with other protein domains at the primary sequence level. In this review, we attempt to categorize the PDZ proteins into three general classes based on their organization properties (fig. 2).

### *Class I: PDZ Domain Proteins with No Intrinsic Enzyme Activities*

This class of PDZ proteins either does not contain any recognizable catalytic domains or has enzyme-like domains, but they lack detectable catalytic activities. This class of PDZ proteins can be further divided into two subclasses.

*Class I-1: PDZ Domain-Only Proteins.* A striking feature of this class of PDZ proteins is that these proteins contain only PDZ domains and no other recognizable protein domains. For example, InaD is composed of 5 PDZ domains, and MUPP1 contains as many as 13 PDZ domains (fig. 2). If each PDZ domain binds to a specific target, one would expect that these multiple PDZ domain proteins will be able to assemble a number of interacting proteins. Therefore, these multiple PDZ domain proteins can function as scaffolds for protein complex assembly in cells. The scaffolding capacity of these multi-PDZ proteins is further expanded by oligomerization of the scaffold protein itself [reviewed in 9, 10]. The best-studied example of PDZ domain-mediated signaling complex assembly is the InaD-mediated photoreception in *Drosophila* [reviewed in 3, 4]. Each of 5 PDZ domains of InaD binds to a specific partner in the fly phototransduction cascade; InaD is, therefore, capable of assembling the multiple signaling components into a 'transducisome' in the rhabdomere of photoreceptor cells [36, 37]. The confinement of signaling proteins onto a single scaffold protein InaD is ideally suited for the remarkably fast (milliseconds) photoreceptive response and adaptation processes in the fly. Mutation of *InaD* leads to mislocalization of a number of proteins (e.g., PKC and PLC), and the mutant fly is nearly blind [3, 4].

*Class I-2: PDZ Domain together with Other Protein Modules.* PDZ domains are found to frequently appear together with other protein domains without detectable enzyme activities. This class of PDZ proteins includes the founding member of the PDZ protein PSD-95. PSD-95 is a prototype of MAGUK (membrane-associated guanylate

kinase) family proteins, which contain one to three PDZ domains followed by an SH3 domain and a guanylate kinase-like (GK) domain. The GK domain of MAGUKs has lost its catalytic activity during evolution, and evolved to be a protein-binding module [38–40]. MAGUKs have been shown to play critical roles in organizing signal complexes in neuronal synapses [5, 8]. In addition to being together with various protein interaction domains such as PTB, LIM, L27 and SH3 domains, PDZ domains also occur in proteins with lipid interaction domains including PH, C2 and PX domains (fig. 2). The cooccurrence of PDZ domains together with other protein-protein and/or protein-lipid interaction domains confers versatile organizational capacities on these scaffold proteins.

### *Class II: PDZ Domain Proteins Containing Enzymatic Activities*

In contrast to the proteins in class I, the class II PDZ proteins contain one or at most two catalytic domains. It seems that various types of enzymatic domains can coexist with PDZ domains. For example, PDZ domains can occur in both protein kinases and phosphatases. Tyrosine phosphatase 1E contains 5 PDZ domains, and the PDZ domains may act as docking sites for various substrates of the enzyme [41, 42]. Neuronal nitric oxide contains a PDZ domain at the N-terminal end of the enzyme, and the function of the PDZ domain is to physically target the enzyme to the NMDA receptor by binding to the PDZ domain of PSD-95 in synapses, thereby the enzyme can be activated in a timely fashion by the NMDA receptor-mediated  $Ca^{2+}$  influx [43].

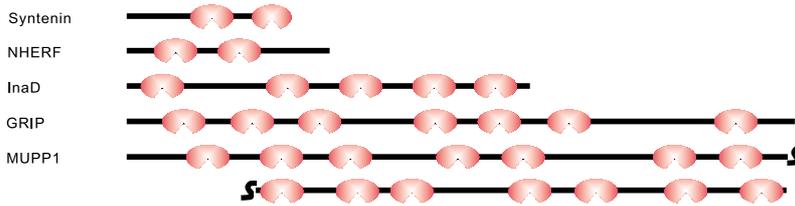
A large number of PDZ proteins in this class contain enzymatic domains that can regulate G protein signaling pathways. For example, various GEF (guanine-nucleotide exchange factor) domains are often associated with PDZ domains. PDZ proteins are often found to contain RGS (regulator of heteromeric G protein signaling) domains. The recently identified PDZ-RGS3 protein is particularly interesting [44]. The PDZ domain of PDZ-RGS3 binds to the C-terminal tail of EphB, and the RGS domain of the protein regulates the GTPase activity of CXCR4, thereby linking the EphB reverse signaling and the G protein couple receptor signaling pathways.

### *Class III: Unconventional PDZ Domain Proteins*

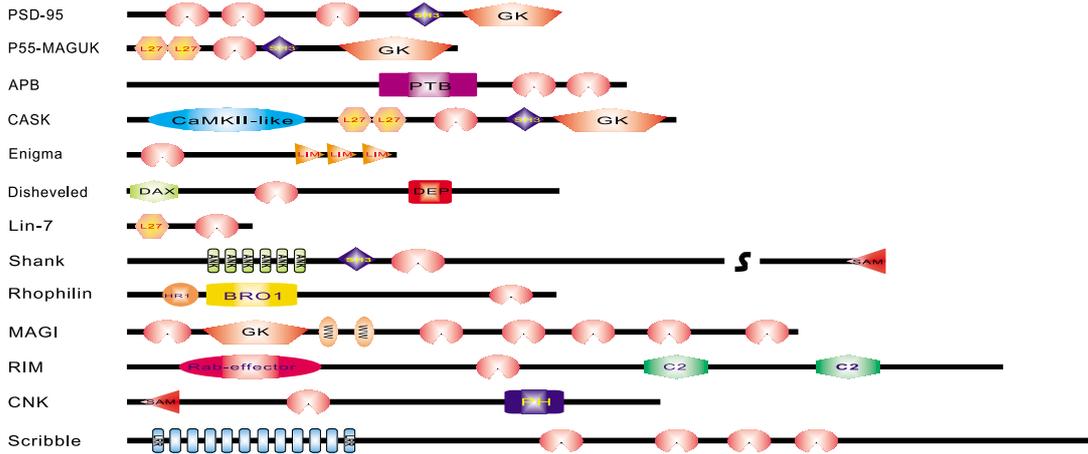
Amino acid sequence analysis identified a distinct family of 'PDZ-like' domain-containing proteins in plants, bacteria as well as metazoans [45]. At the amino acid sequence level, these 'PDZ-like' domains share significantly lower homology with the canonical PDZ do-

Class I: PDZ proteins without intrinsic enzymatic activities

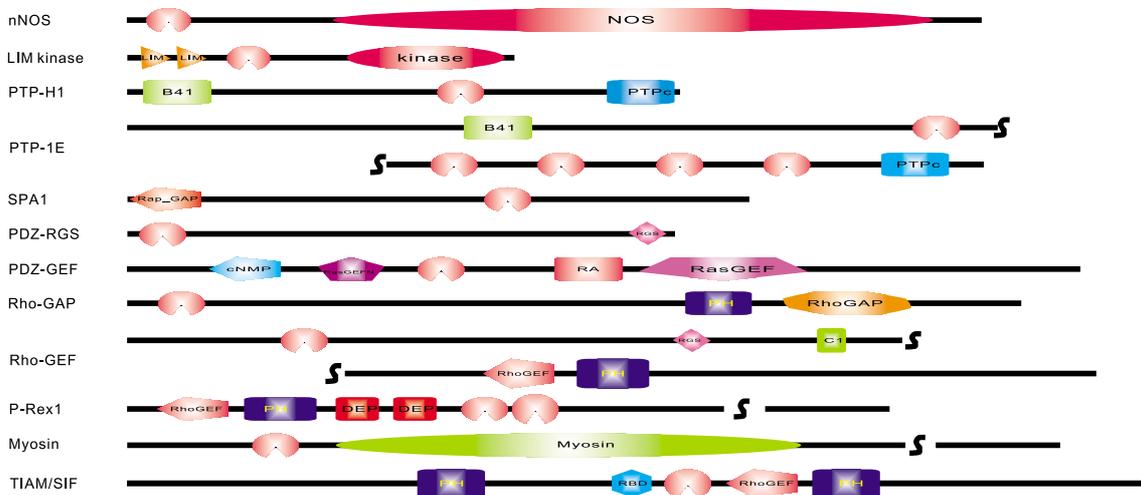
I-1: PDZ domain only proteins



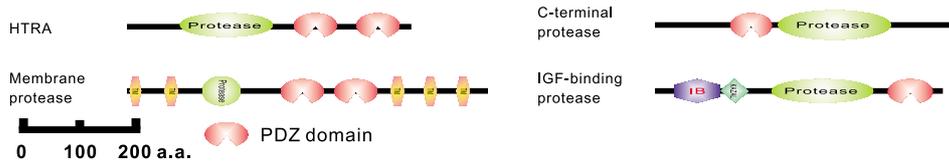
I-2: PDZ domain with other modules



Class II: PDZ proteins that contain enzymatic activities



Class III: Non-conventional PDZ proteins



mains. A signature feature of this family of 'PDZ-like' domain proteins is that the PDZ domains often coexist with various protease domains (fig. 2). Recent crystal structures of three members of this protein family have shown that these 'PDZ-like' domains indeed fold into a three-dimensional structure highly similar to that of the canonical PDZ domains [46–48] (fig. 1c). However, there is a striking difference in their topologies between the protease PDZ domains and the canonical PDZ domains. In the protease PDZ domain, a stretch of amino acid residues at the C-terminal end of the domain forms a  $\beta$ -strand structure and occupies the  $\beta$ A strand position in the canonical PDZ domains (a topology arrangement called circular permutation). The conformation of the  $\alpha$ B/ $\beta$ B groove (the nomenclature of the canonical PDZ domains) of each PDZ domain suggests that the protease PDZ domains can also bind to carboxyl peptides as canonical PDZ domains do [46–48]. In fact, the PDZ2 domain of bacterial HtrA/DegP has an internal peptide fragment bound to the  $\alpha$ B/ $\beta$ B groove [48]. An interesting aspect of the HtrA structures is the direct regulatory roles played by PDZ domains on the catalytic activities of the protease. The PDZ domains can directly interact with the catalytic domain, thereby denying the substrate access to the active site of the enzyme [47, 48]. An attractive working hypothesis is that the PDZ domain acts as the protease substrate

(or regulatory factor) docking site. Binding of the substrates (or regulatory factors) displaces the inhibitory PDZ domains from the active site of the enzyme and leads to the activation of the proteases. It will be interesting to see whether a similar PDZ domain-mediated regulatory strategy is also used in the enzymes containing canonical PDZ domains described in class II PDZ proteins.

#### Novel Aspects of PDZ Structure and Function from Recent Studies

Structural and functional studies in the past few years uncovered several novel features of PDZ domains. In this review, we describe several examples.

##### *PDZ Domains as Lipid-Binding Modules*

A recent study by Zimmermann et al. [49] prompted a rethinking of the current concept of PDZ domains as protein interaction modules. In addition to binding to carboxyl peptides, Zimmermann et al. found that the PDZ domains in a number of proteins including syntenin and CASK can bind to phosphatidylinositol phosphate, and the binding constant between syntenin PDZ1 and lipids is comparable to well-characterized lipid-binding domains such as PH and C2 domains. The authors further showed that the PDZ1 of syntenin selectively binds to phosphatidylinositol 4,5-bisphosphate (PIP<sub>2</sub>). Binding competition experiments using syndecan carboxyl peptide indicated that PIP<sub>2</sub> is likely to occupy the same carboxyl peptide-binding groove of the PDZ domain. The study raises the interesting possibility that PDZ domains contain an intrinsic membrane localization capacity that is independent of binding to receptor tails. It is not known whether lipid binding is a general property of PDZ domains. We tested a number of PDZ domains including PDZ domains from PSD-95 and GRIP, and failed to detect significant binding to various lipids (unpubl. observation).

##### *Tandem PDZ Repeats Function Synergistically in Binding to Targets*

A noticeable feature in multiple PDZ domain proteins is that PDZ domains are often grouped in tandem repeats. For example, PDZ1, 2 of PSD-95 are connected by a short linker of 5 amino acids. The two PDZ domains of syntenin are linked by a 4-residue connecting sequence. PDZ1–3 and PDZ4–6 of GRIP are grouped together. Emerging evidence indicate that these grouped PDZ domains can function synergistically in binding to target proteins [50–52]. Binding of syntenin PDZ2 to carboxyl peptides from

Fig. 2. Domain organization of PDZ proteins. The number of amino acid residues in each protein is indicated in scale by the length of the black line. The detected protein domains (as defined by SMART [55]) are indicated by their names. The abbreviated names of the domains are: SH3 = Src homology 3 domain; GK = guanylate-kinase-like domain; L27 = Lin-2, Lin-7 domain; PTB = protein tyrosine-binding domain; CaMKII-like = calmodulin-dependent kinase II like domain; LIM = zinc-binding domain present in Lin-11, Isl-1, Mec-3; DAX = disheveled and axin-homology domain; DEP = disheveled, Egl-10 and pleckstrin-homology domain; ANK = ankyrin repeats; SAM = sterile alpha motif; HR1 = protein kinase C-related kinase homology region 1 homologues; BRO1 = Bro-1-like domain; WW = domain with 2 conserved Trp residues; RAB effector = C2, protein kinase C-conserved region 2; PH = pleckstrin-homology domain; LRR = leucine-rich repeats; NOS = nitric oxide synthase; B41 = band 4.1 homology domain; PTPc = protein tyrosine phosphatase, catalytic domain; Rap-GAP = rap/ran-GTPase-activating protein homology domain; RGS = regulator of G protein signaling domain; cNMP = cyclic nucleotide-monophosphate-binding domain; RasGEFN = guanine nucleotide exchange factor for Ras-like GTPases N-terminal motif; RA = Ras association domain; RasGEF = Ras-like guanine nucleotide exchange factor; RhoGAP = Rho-like GTPase activator protein; C1 = protein kinase C conserved region 1 domain; RBD = Raf-like Ras-binding domain; TM = transmembrane domain; IB = insulin-like growth factor-binding domain; KAZAL = Kazal-type serine protease inhibitors.

syndecan and ephrin-B1 is positively regulated by PDZ1 [51]. In return, syntenin PDZ1-mediated lipid binding is cooperatively regulated by PDZ2 [49]. PDZ4 of GRIP can directly influence folding of PDZ5, thereby regulating its binding to AMPA receptors [52]. PDZ12 of PSD-95 binds to dimeric ligands with considerably higher affinity than the individual PDZ domains. A recent structural and biochemical study of PSD-95 PDZ12 showed that the two PDZ domains have limited freedom of rotation and their C-terminal peptide-binding grooves are aligned with each other with an orientation preference for binding to pairs of C-termini extending from multimeric membrane proteins [unpubl. observation]. These studies suggested that such tandemly arranged protein-protein interaction modules are not just a simple attachment of 'beads' on a 'string', but may represent functional 'supermodules' spatially organized for coordinated binding to specific targets.

#### *Unusual PDZ Domain-Mediated Interaction*

It is well accepted that PDZ domain-mediated protein target interactions involve the  $\alpha$ B/ $\beta$ B groove of PDZ domains. However, we recently discovered that PDZ7 of GRIP can bind to its target using a surface entirely different from the  $\alpha$ B/ $\beta$ B groove of the domain. GRIP PDZ7 was found to interact with a RasGEF protein called GRASP-1 [53]. The interaction between GRIP and GRASP-1 is rather unusual as GRASP-1 does not contain the carboxyl PDZ-binding motif or a PDZ-like domain. The three-dimensional structure solved by NMR spectroscopy showed that of GRIP PDZ7 contains a large, solvent exposed hydrophobic surface composed of residues from the  $\beta$ E strand, the  $\alpha$ B helix and the loop connecting the two secondary structural elements. It was shown that GRIP PDZ7 interacts with GRASP-1 via this hydrophobic surface [54]. This study uncovered a novel mode of PDZ

domain-mediated protein-protein interaction. It will be interesting to test whether other atypical PDZ domain-mediated interactions also follow a mechanism similar to that observed in the GRIP PDZ7/GRASP-1 complex.

#### Conclusion

Recent studies using genetic and molecular approaches have firmly established that PDZ domain proteins play important roles in assembling protein complexes for signal transduction and organizing large, complicated cellular structures in asymmetric cells such as neurons and epithelial cells. Biochemical and structural studies have provided mechanistic insights into the interaction between PDZ domains and their target proteins. Many questions remain to be answered. For example, how do several hundred PDZ domains control their target-binding specificities inside cells? Do PDZ domains influence each other in the multi-PDZ domain proteins? Where are the PDZ domain protein-mediated signaling complexes assembled/disassembled? Are these signaling complexes dynamically regulated? If yes, how? Another interesting area for future study is to develop specific small molecular weight compounds to intervene between PDZ domain-mediated protein interactions. The use of such compounds may result in potential therapeutic applications regarding various PDZ domain-related human diseases.

#### Acknowledgments

We thank Dr. Andrew Miller for careful reading of the manuscript. Research in MZ's laboratory is partially supported by grants from the Research Grants Council of Hong Kong and the Human Frontier Science Program.

#### References

- Schultz J, Milpetz F, Bork P, Ponting CP: SMART, a simple modular architecture research tool: Identification of signaling domains. *Proc Natl Acad Sci USA* 1998;95:5857-5864.
- Letunic I, Goodstadt L, Dickens NJ, Doerks T, Schultz J, Mott R, Ciccarelli F, Copley RR, Ponting CP, Bork P: Recent improvements to the SMART domain-based sequence annotation resource. *Nucleic Acids Res* 2002;30:242-244.
- Montell C: Visual transduction in *Drosophila*. *Annu Rev Cell Dev Biol* 1999;15:231-268.
- Tsunoda S, Zuker CS: The organization of INAD-signaling complexes by a multivalent PDZ domain protein in *Drosophila* photoreceptor cells ensures sensitivity and speed of signaling. *Cell Calcium* 1999;26:165-171.
- Garner CC, Nash J, Haganir RL: PDZ domains in synapse assembly and signalling. *Trends Cell Biol* 2000;10:274-280.
- Fanning AS, Anderson JM: PDZ domains: Fundamental building blocks in the organization of protein complexes at the plasma membrane. *J Clin Invest* 1999;103:767-772.
- Bilder D: PDZ proteins and polarity: Functions from the fly. *Trends Genet* 2001;17:511-519.
- Craven SE, Bredt DS: PDZ proteins organize synaptic signaling pathways. *Cell* 1998;93:495-498.
- Harris BZ, Lim WA: Mechanism and role of PDZ domains in signaling complex assembly. *J Cell Sci* 2001;114:3219-3231.
- Sheng M, Sala C: PDZ domains and the organization of supramolecular complexes. *Annu Rev Neurosci* 2001;24:1-29.
- Hung AY, Sheng M: PDZ domains: Structural modules for protein complex assembly. *J Biol Chem* 2002;277:5699-5702.
- Morais Cabral JH, Petosa C, Sutcliffe MJ, Raza S, Byron O, Poy F, Marfatia SM, Chishti AH, Liddington RC: Crystal structure of a PDZ domain. *Nature* 1996;382:649-652.
- Doyle DA, Lee A, Lewis J, Kim E, Sheng M, MacKinnon R: Crystal structures of a complexed and peptide-free membrane protein-

- binding domain: Molecular basis of peptide recognition by PDZ. *Cell* 1996;85:1067–1076.
- 14 Daniels DL, Cohen AR, Anderson JM, Brunger AT: Crystal structure of the hCASK PDZ domain reveals the structural basis of class II PDZ domain target recognition. *Nat Struct Biol* 1998;5:317–325.
  - 15 Schultz J, Hoffmuller U, Krause G, Ashurst J, Macias MJ, Schmieder P, Schneider-Mergener J, Oschkinat H: Specific interactions between the syntrophin PDZ domain and voltage-gated sodium channels. *Nat Struct Biol* 1998;5:19–24.
  - 16 Tochio H, Zhang Q, Mandal P, Li M, Zhang M: Solution structure of the extended neuronal nitric oxide synthase PDZ domain complexed with an associated peptide. *Nat Struct Biol* 1999;6:417–421.
  - 17 Hillier BJ, Christopherson KS, Prehoda KE, Brecht DS, Lim WA: Unexpected modes of PDZ domain scaffolding revealed by structure of nNOS-syntrophin complex. *Science* 1999;284:812–815.
  - 18 Tochio H, Hung F, Li M, Brecht DS, Zhang M: Solution structure and backbone dynamics of the second PDZ domain of postsynaptic density-95. *J Mol Biol* 2000;295:225–237.
  - 19 Kozlov G, Gehring K, Ekiel I: Solution structure of the PDZ2 domain from human phosphatase hPTP1E and its interactions with C-terminal peptides from the Fas receptor. *Biochemistry* 2000;39:2572–2580.
  - 20 Karthikeyan S, Leung T, Birrane G, Webster G, Ladias JA: Crystal structure of the PDZ1 domain of human Na<sup>+</sup>/H<sup>+</sup> exchanger regulatory factor provides insights into the mechanism of carboxyl-terminal leucine recognition by class I PDZ domains. *J Mol Biol* 2001;308:963–973.
  - 21 Karthikeyan S, Leung T, Ladias JA: Structural basis of the Na<sup>+</sup>/H<sup>+</sup> exchanger regulatory factor PDZ1 interaction with the carboxyl-terminal region of the cystic fibrosis transmembrane conductance regulator. *J Biol Chem* 2001;276:19683–19686.
  - 22 Kimple ME, Siderovski DP, Sondek J: Functional relevance of the disulfide-linked complex of the N-terminal PDZ domain of InaD with NorpA. *EMBO J* 2001;20:4414–4422.
  - 23 Kozlov G, Banville D, Gehring K, Ekiel I: Solution structure of the PDZ2 domain from cytosolic human phosphatase hPTP1E complexed with a peptide reveals contribution of the beta2-beta3 loop to PDZ domain-ligand interactions. *J Mol Biol* 2002;320:813–820.
  - 24 Karthikeyan S, Leung T, Ladias JA: Structural determinants of the Na<sup>+</sup>/H<sup>+</sup> exchanger regulatory factor interaction with the beta 2 adrenergic and platelet-derived growth factor receptors. *J Biol Chem* 2002;277:18973–18978.
  - 25 Piserchio A, Pellegrini M, Mehta S, Blackman SM, Garcia EP, Marshall J, Mierke DF: The PDZ1 domain of SAP90. Characterization of structure and binding. *J Biol Chem* 2002;277:6967–6973.
  - 26 Songyang Z, Fanning AS, Fu C, Xu J, Marfatia SM, Chishti AH, Crompton A, Chan AC, Anderson JM, Cantley LC: Recognition of unique carboxyl-terminal motifs by distinct PDZ domains. *Science* 1997;275:73–77.
  - 27 Cai C, Coleman SK, Niemi K, Keinonen K: Selective binding of SAP97 to GluR-A AMPA receptor subunit is determined by a novel sequence motif. *J Biol Chem* 2002;277:31484–31490.
  - 28 Gee SH, Sekely SA, Lombardo C, Kurakin A, Froehner SC, Kay BK: Cyclic peptides as non-carboxyl-terminal ligands of syntrophin PDZ domains. *J Biol Chem* 1998;273:21980–21987.
  - 29 Christopherson KS, Hillier BJ, Lim WA, Brecht DS: PSD-95 assembles a ternary complex with the N-methyl-D-aspartic acid receptor and a bivalent neuronal NO synthase PDZ domain. *J Biol Chem* 1999;274:27467–27473.
  - 30 Tochio H, Mok YK, Zhang Q, Kan HM, Brecht DS, Zhang M: Formation of nNOS/PSD-95 PDZ dimer requires a preformed beta-finger structure from the nNOS PDZ domain. *J Mol Biol* 2000;303:359–370.
  - 31 Harris BZ, Hillier BJ, Lim WA: Energetic determinants of internal motif recognition by PDZ domains. *Biochemistry* 2001;40:5921–5930.
  - 32 Dong H, O'Brien RJ, Fung ET, Lanahan AA, Worley PF, Hagan RL: GRIP: A synaptic PDZ domain-containing protein that interacts with AMPA receptors. *Nature* 1997;386:279–284.
  - 33 Srivastava S, Osten P, Vilim FS, Khatri L, Inman G, States B, Daly C, DeSouza S, Abagyan R, Valtchanoff JG, Weinberg RJ, Ziff EB: Novel anchorage of GluR2/3 to the postsynaptic density by the AMPA receptor-binding protein ABP. *Neuron* 1998;21:581–591.
  - 34 Fouassier L, Yun CC, Fitz JG, Doctor RB: Evidence for ezrin-radixin-moesin-binding phosphoprotein 50 (EBP50) self-association through PDZ-PDZ interactions. *J Biol Chem* 2000;275:25039–25045.
  - 35 Lin D, Edwards AS, Fawcett JP, Mbamalu G, Scott JD, Pawson T: A mammalian PAR-3-PAR-6 complex implicated in Cdc42/Rac1 and a PKC signalling and cell polarity. *Nat Cell Biol* 2000;2:540–547.
  - 36 Xu XZ, Choudhury A, Li X, Montell C: Coordination of an array of signaling proteins through homo- and heteromeric interactions between PDZ domains and target proteins. *J Cell Biol* 1998;142:545–555.
  - 37 Tsunoda S, Sierralta J, Sun Y, Bodner R, Suzuki E, Becker A, Socolich M, Zuker CS: A multivalent PDZ-domain protein assembles signaling complexes in a G-protein-coupled cascade. *Nature* 1997;388:243–249.
  - 38 Kim E, Naisbitt S, Hsueh YP, Rao A, Rothschild A, Craig AM, Sheng M: GKAP, a novel synaptic protein that interacts with the guanylate kinase-like domain of the PSD-95/SAP90 family of channel clustering molecules. *J Cell Biol* 1997;136:669–678.
  - 39 Brenman JE, Topinka JR, Cooper EC, McGee AW, Rosen J, Milroy T, Ralston HJ, Brecht DS: Localization of postsynaptic density-93 to dendritic microtubules and interaction with microtubule-associated protein 1A. *J Neurosci* 1998;18:8805–8813.
  - 40 Pak DT, Yang S, Rudolph-Correia S, Kim E, Sheng M: Regulation of dendritic spine morphology by SPAR, a PSD-95-associated Rap-GAP. *Neuron* 2001;31:289–303.
  - 41 Cuppen E, Gerrits H, Pepers B, Wieringa B, Hendriks W: PDZ motifs in PTP-BL and RIL bind to internal protein segments in the LIM domain protein RIL. *Mol Biol Cell* 1998;9:671–683.
  - 42 Palmer A, Zimmer M, Erdmann KS, Eulenburg V, Porthin A, Heumann R, Deutsch U, Klein R: EphrinB phosphorylation and reverse signaling. Regulation by Src kinases and PTP-BL phosphatase. *Mol Cell* 2002;9:725–737.
  - 43 Brenman JE, Chao DS, Gee SH, McGee AW, Craven SE, Santillano DR, Wu Z, Huang F, Xia H, Peters MF, Froehner SC, Brecht DS: Interaction of nitric oxide synthase with the postsynaptic density protein PSD-95 and alpha1-syntrophin mediated by PDZ domains. *Cell* 1996;84:757–767.
  - 44 Lu Q, Sun EE, Klein RS, Flanagan JG: Ephrin-B reverse signaling is mediated by a novel PDZ-RGS protein and selectively inhibits G protein-coupled chemoattraction. *Cell* 2001;105:69–79.
  - 45 Ponting CP: Evidence for PDZ domains in bacteria, yeast, and plants. *Protein Sci* 1997;6:464–468.
  - 46 Liao DI, Qian J, Chisholm DA, Jordan DB, Diner BA: Crystal structures of the photosystem II D1 C-terminal processing protease. *Nat Struct Biol* 2000;7:749–753.
  - 47 Li W, Srinivasula SM, Chai J, Li P, Wu JW, Zhang Z, Alnemri ES, Shi Y: Structural insights into the pro-apoptotic function of mitochondrial serine protease HtrA2/Omi. *Nat Struct Biol* 2002;22:22.
  - 48 Krojer T, Garrido-Franco M, Huber R, Ehrmann M, Clausen T: Crystal structure of DegP (HtrA) reveals a new protease-chaperone machine. *Nature* 2002;416:455–459.
  - 49 Zimmermann P, Meerschaert K, Reekmans G, Leenaerts I, Small JV, Vandekerckhove J, David G, Gettemans J: PIP(2)-PDZ domain binding controls the association of syntenin with the plasma membrane. *Mol Cell* 2002;9:1215–1225.
  - 50 Raghuram V, Mak DO, Foskett JK: Regulation of cystic fibrosis transmembrane conductance regulator single-channel gating by bivalent PDZ-domain-mediated interaction. *Proc Natl Acad Sci USA* 2001;98:1300–1305.
  - 51 Grootjans JJ, Reekmans G, Ceulemans H, David G: Syntenin-syndecan binding requires syndecan-syntenin and the co-operation of both PDZ domains of syntenin. *J Biol Chem* 2000;275:19933–19941.
  - 52 Zhang Q, Fan J-S, Zhang M: Interdomain chaperoning between PSD-95, Dlg, and Zo-1 (PDZ) domains of glutamate receptor-interacting proteins. *J Biol Chem* 2001;276:43216–43220.
  - 53 Ye B, Liao D, Zhang X, Zhang P, Dong H, Hagan RL: GRASP-1: A neuronal RasGEF associated with the AMPA receptor/GRIP complex. *Neuron* 2000;26:603–617.
  - 54 Feng W, Fan J-S, Jiang M, Shi Y-W, Zhang M: PDZ7 of glutamate receptor interacting protein binds to its target via a novel hydrophobic surface area. *J Biol Chem* 2002;277:41140–41146.
  - 55 Schultz J, Copley RR, Doerks T, Ponting CP, Bork P: SMART: A web-based tool for the study of genetically mobile domains. *Nucleic Acids Res* 2000;28:231–234.